

Review

Open Access

Understanding LTP in pain pathways

Jürgen Sandkühler*

Address: Department of Neurophysiology, Center for Brain Research, Medical University of Vienna, Vienna, Austria

Email: Jürgen Sandkühler* - juergen.sandkuehler@meduniwien.ac.at

* Corresponding author

Published: 3 April 2007

Received: 26 March 2007

Molecular Pain 2007, **3**:9 doi:10.1186/1744-8069-3-9

Accepted: 3 April 2007

This article is available from: <http://www.molecularpain.com/content/3/1/9>

© 2007 Sandkühler; licensee BioMed Central Ltd.

This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/2.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Abstract

Long-term potentiation (LTP) at synapses of nociceptive nerve fibres is a proposed cellular mechanism underlying some forms of hyperalgesia. In this review fundamental properties of LTP in nociceptive pathways are described. The following topics are specifically addressed: A concise definition of LTP is given and a differentiation is made between LTP and "central sensitisation". How to (and how not to) measure and how to induce LTP in pain pathways is specified. The signal transduction pathways leading to LTP at C-fibre synapses are highlighted and means of how to preempt and how to reverse LTP are delineated. The potential functional roles of LTP are evaluated at the cellular level and at the behavioural level in experimental animals. Finally, the impact of LTP on the perception of pain in human subjects is discussed.

Background

Long-term potentiation (LTP) is a much studied cellular model of synaptic plasticity. It is generally defined as the long-lasting but not necessarily irreversible increase in synaptic strength [1,2]. At least two different stages of LTP can be distinguished depending upon its duration and the signal transduction pathways involved. Early phase LTP is independent of *de-novo* protein synthesis and lasts for up to three hours. Late phase LTP involves protein synthesis and lasts longer than three hours, up to the life span of an animal and may involve structural changes at synapses [3]. Short-term potentiation of synaptic strength lasts less than half an hour. Synaptic strength is the magnitude of the post-synaptic response (i.e. the post-synaptic potential or the post-synaptic current, but not action potential firing, see below) in response to a pre-synaptic action potential. LTP can be expressed pre- and/or postsynaptically, i.e. synaptic strength can increase if the release of neurotransmitter(s) is enhanced and/or if the postsynaptic effects of the neurotransmitter(s) become stronger [4]. LTP at synapses in hippocampus is the prime model for learning and

memory formation [1]. Recent studies have shown that LTP can also be induced in pain pathways and may contribute to hyperalgesia caused by inflammation, trauma or neuropathy. This review deals with the latter form of LTP.

What is "central sensitisation"?

"Central sensitisation" is used in the literature in at least two mutually exclusive definitions. Some of the authors use "central sensitisation" as an umbrella term for all forms of changes within the central nervous system which ultimately lead to enhanced pain perception. If using this definition of "central sensitisation" one should keep in mind that none of the presently known phenomena in the central nervous system (CNS) which can be observed in experimental or clinical models of hyperalgesia or allodynia has a proven, causative role for the perception of pain. Thus, all presently proposed mechanisms of "central sensitisation" in this definition would have the status of hypotheses. This includes but is not limited to the topic of the present review, the LTP at C-fibre synapses in the superficial spinal dorsal horn.

The task force for taxonomy of the International Association for the Study of Pain (IASP) and many other authors define "central sensitisation" as "an enhanced responsiveness of nociceptive neurons in the CNS to their normal afferent input". This is a very clear-cut definition with little error-proneness. The mechanisms underlying "central sensitisation" in this definition can be well studied experimentally. Nociceptive neurons in the CNS may, however, serve very distinct and also antagonistic functions not all of which are related to the perception of pain. Some nociceptive neurons are excitatory, others are inhibitory. Spinal nociceptive neurons may project to different areas in the brain, with a wide spectrum of functions. Other nociceptive spinal neurons may project segmentally to motoneurons and still other may be interneurons with no known function at all. A remarkable exception are those nociceptive neurons which are located in lamina I of the spinal dorsal horn and which express the NK1 receptor for substance P, many of which project supraspinally. If these neurons are destroyed selectively by a toxin conjugated to substance P, then neither inflammation nor neuropathy leads to full expression of hyperalgesia [5,6]. Interestingly, animal responses to acute noxious stimuli are not altered [5,6]. Thus, these neurons are indispensable for the development of hyperalgesia but not essential for acute pain. Enhanced responsiveness of different nociceptive neurons in the CNS may thus have distinct and perhaps opposing consequences on pain. Thus, "central sensitisation" defined in this way is fundamentally different from the former definition. Some forms of "central sensitisation" as defined by the IASP task force could contribute to hyperalgesia and/or allodynia while other forms may rather lead to the contrary, i.e. stronger feedback inhibition and endogenous pain control. Many of the presently known forms of "central sensitisation" defined in the latter way would be assigned a yet unknown (if any) role for pain perception.

An utter confusion is created if the term "central sensitisation" is used but not defined or when switching from one definition to the other within the same publication, i.e. if both definitions are not clearly distinguished. This would incorrectly imply that any form of enhanced responsiveness of nociceptive neurons in the CNS to their normal afferent input would lead to an enhanced perception of pain. Before we evaluate the evidence suggesting that a prolonged increase in synaptic strength at C-fibres contributes to hyperalgesia we will first review the fundamental properties of LTP in pain pathways.

How to (and how not to) measure LTP

LTP is measured as an increase in mono-synaptically-evoked post-synaptic currents or potentials in response to a single pre-synaptic action potential. LTP is often studied in *in vitro* preparations which allow reliable recordings of

synaptic strength. Whole-cell patch-clamp recording is now the most often used technique. It enables some control over the composition of the intracellular fluid of the post-synaptic neurons which may be advantageous to study post-synaptic mechanisms of LTP. If, however a diffusible mediator is involved and dialysis of the post-synaptic neuron has to be avoided, perforated patch-clamp recordings or intracellular recordings with sharp electrodes can be used. To evaluate LTP at the first synapses in nociceptive pathways, transverse slices with long dorsal roots attached can be prepared from lumbar spinal cord of rats or mice to study mono-synaptic, A δ -fibre or C-fibre evoked excitatory postsynaptic potentials or currents in identified dorsal horn neurons [7,8].

Some aspects of LTP can only be studied in the entire animal with primary afferent nerve fibres and descending pathways from the brain intact. *In vivo* C-fibre-evoked field potentials can be measured in superficial spinal dorsal horn e.g. in response to high intensity electrical stimulation of the sciatic nerve for up to 24 h [9]. These extracellular recorded field potentials reflect summation of post-synaptic, mainly mono-synaptically-evoked currents but not action potential firing [9,10].

Monitoring presynaptic activity at synapses of primary afferent nerve fibres is technically quite demanding. In an attempt to monitor pre-synaptic activity in primary afferents optical recordings techniques have been utilised. Some voltage-sensitive dyes can be anterogradely transported in primary afferents to the central terminals mainly in lamina I [11] and may serve as an indicator for pre-synaptic electrical activity but not for transmitter release.

LTP can not be directly investigated by recording action potential discharges of post-synaptic neurons, as action potential firing not only depends upon synaptic strength but also on membrane excitability and the balance between excitatory and inhibitory input to the neuron. For the same reasons poly-synaptically-evoked responses can generally not be used to study synaptic strength and changes thereof.

How can LTP be induced in pain pathways?

LTP induction by high frequency electrical nerve stimulation

The most frequently used form of conditioning stimulation to induce LTP at synapses in the brain consists of high frequency electrical stimulation (HFS, around 100 Hz) of an input pathway. Likewise, LTP can be induced at spinal synapses of small diameter primary afferents by conditioning high intensity, high frequency burst-like stimulation (typically 100 Hz bursts given several times for 1 s at C-fibre strength) both, *in vitro* and *in vivo*. In spinal cord slice preparations, both, A δ -fibre [7] and C-fibre [8,12]-evoked responses are potentiated by HFS when post-syn-

aptic neurons are mildly depolarized to -70 to -50 mV. The same HFS induces, however, long-term depression (LTD) of A δ -fibre-evoked responses if cells are hyperpolarized to -85 mV suggesting that the polarity of synaptic plasticity is voltage-dependent [7].

Neurons in spinal cord lamina I which express the NK1 receptor play a pivotal role for hyperalgesia in behaving animals [5,6]. Most of these neurons send a projection to supraspinal areas. Interestingly, HFS induces LTP selectively at C-fibre synapses with lamina I neurons which express the NK1 receptor and send a projection to the parabrachial area (Fig. 1). In contrast, HFS fails to induce LTP at synapses with neurons which express the NK1 receptor and send a projection to the periaqueductal grey or at synapses with neurons which do not express the NK1 receptor and which have no identified supraspinal projection [8,12].

HFS at C-fibre intensity of sciatic nerve fibre afferents induces LTP of C-fibre, but not A β -fibre-evoked field potentials in superficial spinal dorsal horn of adult, deeply anaesthetized rats [9,13,14]. In contrast, conditioning HFS at A-fibre intensity fails to induce LTP of either A- or C-fibre-evoked field potentials in intact animals. In spinalised animals, conditioning HFS at A δ -fibre intensity induces, however, LTP of C-fibre-evoked field potentials [15].

LTP induced by low frequency electrical nerve stimulation

For most of the C-fibre afferents it is not typical to discharge at rates as high as 100 imp. s⁻¹. Some C-fibres may, however, discharge at these high rates but only for short periods of time, e.g. at the beginning of a noxious mechanical stimulus [16]. Many C-fibres discharge at considerably lower rates, around 1–10 imp. s⁻¹, e.g. in response to an inflammation or an injury [17]. Conditioning stimulation within this lower frequency band is successfully used to induce LTP at C-fibre synapses. In a spinal cord-dorsal root slice preparation conditioning electrical low frequency stimulation (LFS 2 Hz for 2–3 min, C-fibre strength) of dorsal root afferents induces LTP selectively at C-fibre synapses with lamina I neurons that express the NK1 receptor and project to the periaqueductal grey (Fig. 1) [12]. C-fibre synapses with lamina I neurons which express the NK1 receptor and project to the parabrachial area or with no identified supraspinal projection are, in contrast, not potentiated by LFS [12]. Thus, the pattern and the frequency of discharges in C-fibres determine which synapses at the origin of different ascending pain pathways are potentiated.

Pairing strong post-synaptic depolarization (to +30 mV) with low frequency (2 Hz) stimulation of afferent nerve fibres also leads to a robust LTP in superficial spinal dorsal

horn neurons *in vitro* [18]. In spinal cord slices from neonatal rats field potentials evoked by electrical stimulation in the tract of Lissauer are potentiated by repetitive burst-like stimulation at 10 Hz [19].

In deeply anaesthetized adult rats with their spinal cords left intact LFS (at 2 Hz for 2–3 min) of sciatic nerve fibres at C-fibre intensity but not at A δ -fibre intensity also triggers LTP of C-fibre-evoked potentials (Fig. 1) [12].

In conclusion, HFS and LFS may have fundamentally different effects on LTP induction at different C-fibre synapses. This finding is in line with previous reports also illustrating that the frequency of afferent barrage in C-fibres may have qualitatively different effects in spinal cord. For example, brain-derived neurotrophic factor is released from primary afferents in spinal cord slices in an activity-dependent manner by HFS at 100 Hz but not by 1 Hz LFS of primary afferent nerve fibres [20]. Furthermore, in spinal cord slices from mice HFS (100 Hz) of primary afferent nerve fibres at C-fibre intensity, but not LFS (900 pulses at 1 Hz) selectively induces phosphorylation of extracellular receptor-activated MAP Kinases (ERK1/2) in spinal dorsal horn lamina I [21].

Natural noxious stimulation induces LTP in pain pathways

At synapses in the brain LTP induction requires synchronous, high-frequency pre-synaptic activity or pairing of low-level pre-synaptic activity with strong post-synaptic depolarization. At least some of the C-fibre synapses are apparently unique in that LTP can be induced by LFS and by natural, low or high frequency, asynchronous and irregular discharge patterns in sensory nerve fibres. In animals with spinal cord and descending pathways intact, intraplantar, s.c. injections of capsaicin (100 μ l, 1%) or formalin (100 μ l, 5%) induce slowly rising LTP (Fig. 1) [12].

Some forms of low level afferent input can induce LTP only if descending, presumably inhibitory pathways are interrupted or weakened. Noxious radiant heating of the skin at a hind paw induces LTP in spinalised animals but not in animals with spinal cord intact [22]. Likewise, repetitive, noxious squeezing of the skin or the sciatic nerve induces LTP of C-fibre-evoked field potentials in spinalised rats only [22]. These findings indicate that endogenous antinociceptive systems not only raise thresholds for nociception but also those for the induction of LTP.

Pharmacological induction of LTP

At C-fibre synapses LTP can also be induced in the absence of any pre-synaptic activity. Spinal application of a dopamine 1/dopamine 5 receptor agonist (SKF 38393) *in vivo* induces a slowly developing LTP of C-fibre-evoked

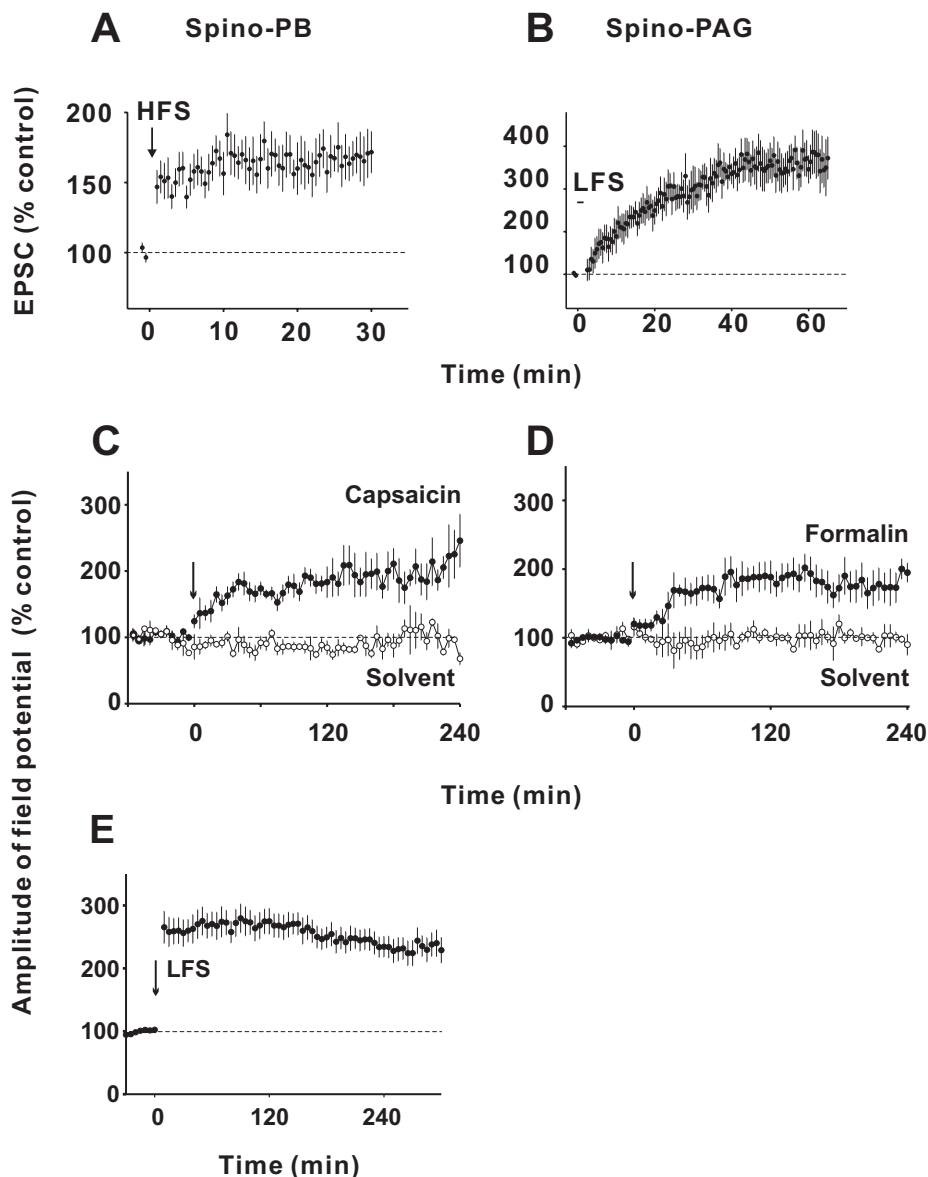


Figure 1

A, B show contrasting forms of LTP expressed in distinct groups of spinal lamina I projection neurons *in vitro*. Time courses of mean amplitudes (\pm SEM) of C-fibre-evoked EPSCs in lamina I neurons with a projection to the parabrachial area (PB, $n = 8$) or the periaqueductal grey (PAG, $n = 7$). Conditioning HFS induced LTP in all spino-PB neurons tested (**A**) but was ineffective in spino-PAG neurons (data not shown). Conditioning LFS induced LTP in all 18 spino-PAG neurons tested (**B**) but was never effective in seven spino-PB neurons (data not shown). **C-E** LTP can be induced by natural, low frequency afferent barrage evoked by inflammation of peripheral tissue *in vivo*. Mean time courses of C-fibre-evoked field potentials recorded extracellularly in superficial spinal dorsal horn in response to electrical stimulation of left sciatic nerve of deeply anesthetized adult rats with spinal cords and afferent nerves intact. Subcutaneous injections of transient receptor potential vanilloid 1 channel agonist capsaicin (1%, 100 μ l, $n = 5$, **C**) or formalin (5%, 100 μ l, $n = 6$, **D**) into the glabrous skin at the ipsilateral hind paw, within the innervation territory of the sciatic nerve at time zero (arrows) induced LTP (closed circles), while injections of the respective solvents (open circles) had no effects ($n = 3$ in each group). Conditioning electrical LFS (2 Hz, 2 min at C-fibre intensity) of sciatic nerve at time zero (arrow) induced LTP ($n = 28$, **E**) which was prevented by NMDA receptor antagonist MK-801 (3 mg kg⁻¹, i.v.-infusion over 30 min, data not shown). Modified from [12].

field potentials which lasts for at least 10 h and which is blocked by the dopamine 1/dopamine 5 antagonist SCH 23390 [23]. In spinalised, deeply anaesthetized, adult rats, superfusions of spinal cord segments with NMDA (10 μ M), substance P (10 μ M) or neurokinin A (1 μ M) are all sufficient to induce LTP of C-fibre-evoked field potentials [24]. With spinal cord and descending (inhibitory) pathways intact spinal applications of NMDA (1–100 μ M), substance P (1–100 μ M) or neurokinin A (1 or 10 μ M) fail, however, to induce LTP of C-fibre-evoked field potentials [24].

LTP of A-fibre-evoked responses

A-fibre-evoked spinal field potentials are depressed by conditioning 50 Hz stimulation of sciatic nerve fibres. After GABA_A receptor antagonist bicuculline (1 mg/kg i.p.) the same conditioning stimulus now produces LTP rather than LTD [25]. Similarly, 50 Hz conditioning stimulation produces short lasting potentiation followed by LTD in control animals but LTP in animals with a CCI of sciatic nerve [26]. Topical application of muscimol (10 μ g), a GABA_A receptor agonist to spinal cord prevents tetanus-induced LTP of A-fibre-evoked field potentials in animals with a CCI [27]. This again suggests that the polarity of synaptic plasticity is context-sensitive and not solely dominated by the type of afferent input.

Signal transduction pathways leading to LTP at C-fibre synapses

In principle, LTP can be induced and/or expressed by pre-synaptic [28] or by post-synaptic [29,30] mechanisms or by any combination thereof. At present, there is clear evidence for a post-synaptic, Ca²⁺-dependent form of LTP induction in spinal cord lamina I neurons. Induction of LTP at C-fibre-synapses requires co-activation of NK1 and NK2 receptors [9], opening of ionotropic glutamate receptors of the NMDA type [8,12,13], opening of T-type voltage-gated calcium channels [8,12], and activation of group I but not group II or III metabotropic glutamate receptors [31]. Activation of NK1 receptors by substance P may directly enhance single NMDA channel opening [32] and NMDA receptor mediated currents in lamina I neurons [8] and all this may lead to substantial rise in post-synaptic [Ca²⁺]_i (Fig. 2). It is presently unknown if Ca²⁺ influx through Ca²⁺-permeable α -amino-3-hydroxy-5-methyl-4-isoxazolepropionic acid (AMPA) receptors is required for LTP induction in pain pathways. Some indirect evidence suggests, however, that this might be the case [21,33].

In any case, a rise in post-synaptic [Ca²⁺]_i is essential for LTP induction and the magnitude in [Ca²⁺]_i rise is linearly correlated with the magnitude of LTP *in vitro* [8]. Recent data demonstrate that LTP-inducing stimuli cause substantial rise in [Ca²⁺]_i in lamina I neurons not only in slice

preparations, but also in intact animals [12]. Not surprisingly therefore signal transduction involves Ca²⁺-dependent pathways including activation of protein kinase C, calcium-calmodulin-dependent protein kinase II (CaMKII), protein kinase A (PKA) phospholipase C (PLC), inositoltriphosphate-3 (IP₃) receptors, nitric oxide synthase (NOS) and members of the mitogen-activated protein kinase family (MAPK), including extracellular signal-regulated kinase (ERK) (Fig. 2) [8,12,18,34-36].

When assessed with voltage-sensitive dyes the pre-synaptic facilitation of electrical activity in primary afferents after LTP-inducing stimuli is partially sensitive to iNOS inhibitor (AMT), a blocker of glial cell metabolisms (MFA), and an mGluR group I antagonist (LY367385) [11].

Inhibition of protein synthesis in spinal cord by either cycloheximide or anisomycin selectively inhibits the maintenance of the late-phase of spinal LTP but does not affect either LTP induction or baseline responses of C-fibre evoked field potentials [37].

Importantly, the very same signal transduction pathways are required for full expression of hyperalgesia in animal models of inflammatory and neuropathic pain e.g. [38-41].

How to pre-empt LTP induction in pain pathways

LTP induction can be prevented by blockade of any of the above mentioned essential elements of signal transduction for LTP. In mature rats deep (surgical) level of anaesthesia with either urethane, isoflurane or sevoflurane is, however, insufficient to pre-empt LTP induction of C-fibre-evoked field potentials [42]. LTP is prevented by low dose intravenous infusion of μ -opioid receptor agonist fentanyl [42]. Similarly, LTP of spinal field potentials elicited by stimulation in the tract of Lissauer in spinal cord slices is blocked by DAMGO, a more specific agonist at these receptors [19]. Activation of spinal α_2 -adrenoreceptors by clonidine [43] or spinal application of the benzodiazepine diazepam [44] also prevents LTP induction *in vivo*.

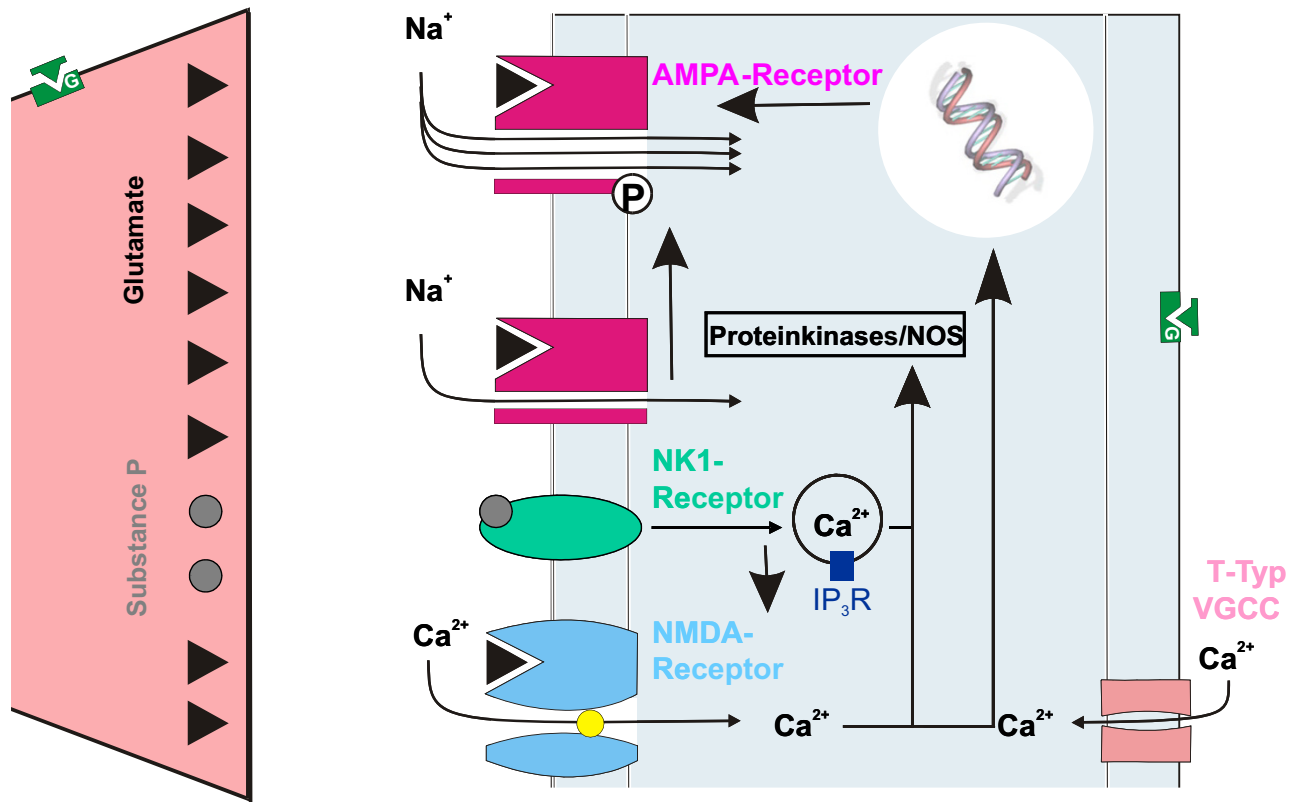
Functional blockade of glial cells by i.t. administration of fluorocitrate changes the polarity of HFS induced synaptic plasticity. When HFS is given 1 h, but not 3 h after fluorocitrate LTD but no LTP of C-fibre-evoked field potentials is induced [14].

LTP can be reversed

LTP of C-fibre-evoked field potentials can be reversed by brief, high frequency conditioning electrical stimulation of sciatic nerve fibres at A δ -fibre intensity [15]. Reversal of LTP by A δ -fibre stimulation is time-dependent and effec-

C-fibre terminal

Lamina I projection neuron

**Figure 2**

Potential mechanisms of potentiation, prevention and de-potentiation at synapses between C-fibres and spinal cord projection neurons. Conditioning electrical nerve stimulation or natural noxious stimulation triggers release of glutamate and substance P which causes opening of NMDA receptor channels and T-type voltage-gated Ca²⁺ channel and Ca²⁺ release from intracellular stores. This activates Ca²⁺-dependent signal transduction pathways including protein kinases and transcription factors. Synaptic strength is probably increased by phosphorylation of synaptic proteins including AMPA receptor channels [55], altered trafficking of synaptic proteins, e.g. increased insertion of AMPA receptors into the sub-synaptic membrane [56] and de-novo protein synthesis. According to this model, LTP can be prevented if release of glutamate and/or substance P is inhibited, for example by activation of pre-synaptic, G-protein-coupled μ -opioid receptors, or if opening of voltage sensitive and Ca²⁺ permeable ion channels is blocked, e.g. via postsynaptic inhibition by an opioid. Depotentiation could result from de-phosphorylation of synaptic proteins, changes in receptors trafficking and degradation of synaptic proteins.

tive only when applied 15 or 60 min but not 3 h after LTP induction [45].

Spinal application of either, NK1 or NK2 receptor antagonists one to three hours after HFS, i.e., after LTP is established, does not affect maintenance of LTP [9], suggesting that activation of these receptors, which are required for the induction of LTP are not essential for its maintenance.

What is the functional role of LTP in pain pathways?

Modulation of synaptic strength is a powerful mechanism to control signal flow in selected pathways. A typical con-

sequence of LTP at excitatory synapses would be an increase in action potential firing of the same and perhaps also of downstream neurons in response to a given stimulus. And indeed, LTP-inducing conditioning stimuli have been found to facilitate action potential firing of multireceptive neurons in deep dorsal horn e.g. [46-48]. This is likely due to LTP at the first synapse in the nociceptive pathway but other mechanisms of facilitation should not be excluded. Action potential firing would also be enhanced if membrane excitability is increased, i.e. the thresholds for action potential firing are lowered, if inhibition is less effective or if inhibition is even reversed and

becomes excitatory e.g. due to a reversal of the anion gradient in the post-synaptic neuron [49,50].

HFS of sciatic nerve fibres which induces LTP at synapses of C-fibres in spinal cord has behavioural consequences in rats and causes thermal hyperalgesia at the ipsilateral hind paw for six days [34]. This suggests that LTP at C-fibre synapses has an impact on nociceptive behaviour.

Perceptual correlates of LTP in pain pathways in human subjects

An indispensable proof for any proposed mechanism of hyperalgesia is an appropriate correlate in the human. And indeed, conditioning HFS of cutaneous peptidergic afferents in humans cause increased pain perception in response to electrical test stimuli applied through the same stimulation electrode [51]. Noxious stimulation with punctate mechanical probes in skin adjacent to the HFS conditioning skin site uncovers a marked (2–3 fold) increase in pain sensitivity, i.e. secondary hyperalgesia [51]. Touching the skin around the conditioning stimulation electrode with a soft cotton wisp evokes pain only after HFS. Thus, HFS also induces secondary mechanical allodynia. Hyperalgesia at the conditioned site but not secondary hyperalgesia or allodynia at adjacent skin areas is prevented by pre-treatment with ketamine [52], a clinically used substance which, among other effects, also blocks NMDA receptors.

Interestingly, all thermal modalities comprising cold and warm detection thresholds, cold and heat pain thresholds as well as pain summation (perceptual "wind-up") remain unaltered after conditioning HFS of peptidergic skin nerve fibres [53].

When verbal pain descriptors are used to evaluate pain in addition to its perceived intensity after HFS, a significant long-term increase in scores for sensory but not for affective descriptors of pain is detected [54]. Within the sensory descriptors, those describing superficial pain, those for heat pain and those for sharp mechanical pain are all potentiated. The authors conclude that brief painful stimuli rarely have a strong affective component and that perceived pain after HFS exhibits predominantly a potentiation of the C-fibre-mediated percepts hot and burning [54].

In humans subjects conditioning LFS causes also an increased pain sensitivity in the area around the LFS conditioned skin site but a depression of pain evoked by stimulation through the same electrode [51].

Conclusion

LTP at synapses between primary afferent C-fibres and a group of nociceptive neurons in spinal cord lamina I

which express the NK1 receptor for substance P is a potential mechanism underlying some forms of pain amplification in behaving animals and perhaps human subjects. Both, LTP and hyperalgesia involve the same essential elements, i.e. primary afferent C-fibres and lamina I neurons which express the NK1 receptor. Further, induction protocols, pharmacological profile and signal transduction pathways are virtually identical.

Abbreviations

AMPA α -amino-3-hydroxy-5-methyl-4-isoxazolepropionic acid

CaMKII Calcium-calmodulin-dependent protein kinase II

CNS Central nervous system

HFS High frequency stimulation

IASP International Association for the Study of Pain

IP₃ Inositol 1,4,5-trisphosphate

LFS Low frequency stimulation

LTP Long-term potentiation

MAPK Mitogen-activated protein kinase

NK1 Neurokinin 1

NK2 Neurokinin 2

NMDA N-methyl-D-aspartate

NOS Nitric oxide synthase

PKA Protein kinase A

PKC Protein kinase C

PLC Phospholipase C

Competing interests

The author declares that he has no competing interests.

Acknowledgements

I wish to thank Professor Ruth Ruscheweyh for helpful comments on an earlier version of the manuscript. The authors work is supported by grants from the Austria Science Fund (FWF) grant # P18129 and # P19367.

References

1. Bliss TVP, Collingridge GL: **A synaptic model of memory: long-term potentiation in the hippocampus.** *Nature* 1993, **361**:31-39.
2. Malenka RC, Bear MF: **LTP and LTD: an embarrassment of riches.** *Neuron* 2004, **44**:5-21.

3. Bailey CH, Kandel ER, Si K: **The persistence of long-term memory: a molecular approach to self-sustaining changes in learning-induced synaptic growth.** *Neuron* 2004, **44**:49-57.
4. Lisman J, Raghavachari S: **A unified model of the presynaptic and postsynaptic changes during LTP at CA1 synapses.** *Sci STKE* 2006, **2006**:1-15.
5. Mantyh PW, Rogers SD, Honoré P, Allen BJ, Ghilardi JR, Li J, et al.: **Inhibition of hyperalgesia by ablation of lamina I spinal neurons expressing the substance P receptor.** *Science* 1997, **278**:275-279.
6. Nichols ML, Allen BJ, Rogers SD, Ghilardi JR, Honoré P, Luger NM, et al.: **Transmission of chronic nociception by spinal neurons expressing the substance P receptor.** *Science* 1999, **286**:1558-1561.
7. Randic M, Jiang MC, Cerne R: **Long-term potentiation and long-term depression of primary afferent neurotransmission in the rat spinal cord.** *J Neurosci* 1993, **13**:5228-5241.
8. Ikeda H, Heinke B, Ruscheweyh R, Sandkühler J: **Synaptic plasticity in spinal lamina I projection neurons that mediate hyperalgesia.** *Science* 2003, **299**:1237-1240.
9. Liu XG, Sandkühler J: **Characterization of long-term potentiation of C-fiber-evoked potentials in spinal dorsal horn of adult rat: essential role of NK1 and NK2 receptors.** *J Neurophysiol* 1997, **78**:1973-1982.
10. Schouenborg J: **Functional and topographical properties of field potentials evoked in rat dorsal horn by cutaneous C-fiber stimulation.** *J Physiol* 1984, **356**:169-192.
11. Ikeda H, Murase K: **Glial nitric oxide-mediated long-term presynaptic facilitation revealed by optical imaging in rat spinal dorsal horn.** *J Neurosci* 2004, **24**:9888-9896.
12. Ikeda H, Stark J, Fischer H, Wagner M, Drdla R, Jäger T, et al.: **Synaptic amplifier of inflammatory pain in the spinal dorsal horn.** *Science* 2006, **312**:1659-1662.
13. Liu XG, Sandkühler J: **Long-term potentiation of C-fiber-evoked potentials in the rat spinal dorsal horn is prevented by spinal N-methyl-D-aspartic acid receptor blockage.** *Neurosci Lett* 1995, **191**:43-46.
14. Ma J-Y, Zhao Z-Q: **The involvement of glia in long-term plasticity in the spinal dorsal horn of the rat.** *Neuroreport* 2002, **13**:1781-1784.
15. Liu XG, Morton CR, Azkue JJ, Zimmermann M, Sandkühler J: **Long-term depression of C-fiber-evoked spinal field potentials by stimulation of primary afferent Aδ-fibers in the adult rat.** *Eur J Neurosci* 1998, **10**:3069-3075.
16. Handwerker HO, Anton F, Reeh PW: **Discharge patterns of afferent cutaneous nerve fibers from the rat's tail during prolonged noxious mechanical stimulation.** *Exp Brain Res* 1987, **65**:493-504.
17. Puig S, Sorkin LS: **Formalin-evoked activity in identified primary afferent fibers: systemic lidocaine suppresses phase-2 activity.** *Pain* 1996, **64**:345-355.
18. Wei F, Vadakkan KI, Toyoda H, Wu L-J, Zhao M-G, Xu H, et al.: **Calcium calmodulin-stimulated adenylyl cyclases contribute to activation of extracellular signal-regulated kinase in spinal dorsal horn neurons in adult rats and mice.** *J Neurosci* 2006, **26**:851-861.
19. Terman GW, Eastman CL, Chavkin C: **Mu opiates inhibit long-term potentiation induction in the spinal cord slice.** *J Neurophysiol* 2001, **85**:485-494.
20. Lever IJ, Bradbury EJ, Cunningham JR, Adelson DW, Jones MG, McMahon SB, et al.: **Brain-derived neurotrophic factor is released in the dorsal horn by distinctive patterns of afferent fiber stimulation.** *J Neurosci* 2001, **21**:4469-4477.
21. Hartmann B, Ahmadi S, Heppenstall PA, Lewin GR, Schott C, Borhardt T, et al.: **The AMPA receptor subunits GluR-A and GluR-B reciprocally modulate spinal synaptic plasticity and inflammatory pain.** *Neuron* 2004, **44**:637-650.
22. Sandkühler J, Liu X: **Induction of long-term potentiation at spinal synapses by noxious stimulation or nerve injury.** *Eur J Neurosci* 1998, **10**:2476-2480.
23. Yang H-W, Zhou L-J, Hu N-W, Xin W-J, Liu X-G: **Activation of spinal D1/D5 receptors induces late phase LTP of c-fiber evoked field potentials in rat spinal dorsal horn.** *J Neurophysiol* 2005, **94**:961-967.
24. Liu XG, Sandkühler J: **Activation of spinal N-methyl-D-aspartate or neurokinin receptors induces long-term potentiation of spinal C-fiber-evoked potentials.** *Neuroscience* 1998, **86**:1209-1216.
25. Miletic G, Miletic V: **Contribution of GABA-A receptors to metaplasticity in the spinal dorsal horn.** *Pain* 2001, **90**:157-162.
26. Miletic G, Miletic V: **Long-term changes in sciatic-evoked A-fiber dorsal horn field potentials accompany loose ligation of the sciatic nerve in rats.** *Pain* 2000, **84**:353-359.
27. Miletic G, Draganic P, Pankratz MT, Miletic V: **Muscimol prevents long-lasting potentiation of dorsal horn field potentials in rats with chronic constriction injury exhibiting decreased levels of the GABA transporter GAT-1.** *Pain* 2003, **105**:347-353.
28. Mellor J, Nicoll RA, Schmitz D: **Mediation of hippocampal mossy fiber long-term potentiation by presynaptic I_h channels.** *Science* 2002, **295**:143-147.
29. Nicoll RA: **Expression mechanisms underlying long-term potentiation: a postsynaptic view.** *Philos Trans R Soc Lond B Biol Sci* 2003, **358**:721-726.
30. Malinow R: **AMPA receptor trafficking and long-term potentiation.** *Philos Trans R Soc Lond B Biol Sci* 2003, **358**:707-714.
31. Azkue JJ, Liu XG, Zimmermann M, Sandkühler J: **Induction of long-term potentiation of C fiber-evoked spinal field potentials requires recruitment of group I, but not group II/III metabotropic glutamate receptors.** *Pain* 2003, **106**:373-379.
32. Lieberman DN, Mody I: **Substance P enhances NMDA channel function in hippocampal dentate gyrus granule cells.** *J Neurophysiol* 1998, **80**:113-119.
33. Tong C-K, MacDermott AB: **Both Ca²⁺-permeable and -impermeable AMPA receptors contribute to primary synaptic drive onto rat dorsal horn neurons.** *J Physiol* 2006, **575**:133-144.
34. Zhang X-C, Zhang Y-Q, Zhao Z-Q: **Involvement of nitric oxide in long-term potentiation of spinal nociceptive responses in rats.** *Neuroreport* 2005, **16**:1197-1201.
35. Xin W-J, Gong Q-J, Xu J-T, Yang H-W, Zang Y, Zhang T, et al.: **Role of phosphorylation of ERK in induction and maintenance of LTP of the C-fiber evoked field potentials in spinal dorsal horn.** *J Neurosci Res* 2006, **84**:934-943.
36. Yang H-W, Hu X-D, Zhang H-M, Xin W-J, Li M-T, Zhang T, et al.: **Roles of CaMKII, PKA and PKC in the induction and maintenance of LTP of C-fiber evoked field potentials in rat spinal dorsal horn.** *J Neurophysiol* 2004, **91**:1122-1133.
37. Hu N-W, Zhang H-M, Hu X-D, Li M-T, Zhang T, Zhou L-J, et al.: **Protein synthesis inhibition blocks the late-phase LTP of C-fiber evoked field potentials in rat spinal dorsal horn.** *J Neurophysiol* 2003, **89**:2354-2359.
38. Sandkühler J: **Learning and memory in pain pathways.** *Pain* 2000, **88**:113-118.
39. Willis WD: **Role of neurotransmitters in sensitization of pain responses.** *Ann N Y Acad Sci* 2001, **933**:142-156.
40. Meller ST, Gebhart GF: **Nitric oxide (NO) and nociceptive processing in the spinal cord.** *Pain* 1993, **52**:127-136.
41. Petersen-Zeitl KR, Basbaum AI: **Second messengers, the substantia gelatinosa and injury-induced persistent pain.** *Pain* 1999:55-12.
42. Benrath J, Brechtel C, Martin E, Sandkühler J: **Low doses of fentanyl block central sensitization in the rat spinal cord in vivo.** *Anesthesiology* 2004, **100**:1545-1551.
43. Ge Y-X, Xin W-J, Hu N-W, Zhang T, Xu J-T, Liu X-G: **Clonidine depresses LTP of C-fiber evoked field potentials in spinal dorsal horn via NO-cGMP pathway.** *Brain Res* 2006, **1118**:58-65.
44. Hu X-D, Ge Y-X, Hu N-W, Zhang H-M, Zhou L-J, Zhang T, et al.: **Diazepam inhibits the induction and maintenance of LTP of C-fiber evoked field potentials in spinal dorsal horn of rats.** *Neuropharmacology* 2006, **50**:238-244.
45. Zhang H-M, Qi Y-J, Xiang X-Y, Zhang T, Liu X-G: **Time-dependent plasticity of synaptic transmission produced by long-term potentiation of C-fiber evoked field potentials in rat spinal dorsal horn.** *Neurosci Lett* 2001, **315**:81-84.
46. Afrah AW, Fiskå A, Gjerstad J, Gustafsson H, Tjølsen A, Olgart L, et al.: **Spinal substance P release in vivo during the induction of long-term potentiation in dorsal horn neurons.** *Pain* 2002, **96**:49-55.
47. Rygh LJ, Suzuki R, Rahman W, Wong Y, Vonsy JL, Sandhu H, et al.: **Local and descending circuits regulate long-term potentiation and zif268 expression in spinal neurons.** *Eur J Neurosci* 2006, **24**:761-772.

48. Vikman KS, Duggan AW, Siddall PJ: **Increased ability to induce long-term potentiation of spinal dorsal horn neurones in monoarthritic rats.** *Brain Res* 2003, **990**:51-57.
49. Coull JAM, Boudreau D, Bachand K, Prescott SA, Nault F, Sik A, et al.: **Trans-synaptic shift in anion gradient in spinal lamina I neurons as a mechanism of neuropathic pain.** *Nature* 2003, **424**:938-942.
50. Coull JAM, Beggs S, Boudreau D, Boivin D, Tsuda M, Inoue K, et al.: **BDNF from microglia causes the shift in neuronal anion gradient underlying neuropathic pain.** *Nature* 2005, **438**:1017-1021.
51. Klein T, Magerl W, Hopf H-C, Sandkühler J, Treede R-D: **Perceptual correlates of nociceptive long-term potentiation and long-term depression in humans.** *J Neurosci* 2004, **24**:964-971.
52. Klein T, Magerl W, Nickel U, Hopf H-C, Sandkühler J, Treede R-D: **Effects of the NMDA-receptor antagonist ketamine on perceptual correlates of long-term potentiation within the nociceptive system.** *Neuropharmacology* 2006, **52**:655-661.
53. Lang S, Klein T, Magerl W, Treede R-D: **Modality-specific sensory changes in humans after the induction of long-term potentiation (LTP) in cutaneous nociceptive pathways.** *Pain* 2007, **128**:254-263.
54. Hansen N, Klein T, Magerl W, Treede R-D: **Psychophysical evidence for long-term potentiation of C-fiber and A δ -fiber pathways in humans by analysis of pain descriptors.** *J Neurophysiol* 2007, **97**:2559-2563.
55. Lee H-K, Takamiya K, Han J-S, Man H, Kim C-H, Rumbaugh G, et al.: **Phosphorylation of the AMPA receptor GluR1 subunit is required for synaptic plasticity and retention of spatial memory.** *Cell* 2003, **112**:631-643.
56. Boehm J, Kang M-G, Johnson RC, Esteban J, Huganir RL, Malinow R: **Synaptic Incorporation of AMPA Receptors during LTP Is Controlled by a PKC Phosphorylation Site on GluR1.** *Neuron* 2006, **51**:213-225.

Publish with **BioMed Central** and every scientist can read your work free of charge

"BioMed Central will be the most significant development for disseminating the results of biomedical research in our lifetime."

Sir Paul Nurse, Cancer Research UK

Your research papers will be:

- available free of charge to the entire biomedical community
- peer reviewed and published immediately upon acceptance
- cited in PubMed and archived on PubMed Central
- yours — you keep the copyright

Submit your manuscript here:
http://www.biomedcentral.com/info/publishing_adv.asp

