

Research

Open Access

## Intracerebroventricular administration of *N*-acetylaspartylglutamate (NAAG) peptidase inhibitors is analgesic in inflammatory pain

Tatsuo Yamamoto<sup>1,5</sup>, Alan Kozikowski<sup>2</sup>, Jia Zhou<sup>3</sup> and Joseph H Neale<sup>\*4</sup>

Address: <sup>1</sup>Department of Anaesthesiology, Graduate School of Medicine, Chiba University, 1-8-1 Inohana, Chuo-ku, Chiba-shi, Chiba 260-8670, Japan, <sup>2</sup>Drug Discovery Program, Department of Medicinal Chemistry and Pharmacognosy, University of Illinois at Chicago, Chicago, Illinois 60612, USA, <sup>3</sup>PsychoGenics Inc., Tarrytown, NY 10591, USA, <sup>4</sup>Department of Biology, Georgetown University, Washington, D.C., 20057, USA and <sup>5</sup>Department of Anesthesiology at Kumamoto University, Kumamoto, Japan

Email: Tatsuo Yamamoto - yamamoto@fc.kuh.kumamoto-u.ac.jp; Alan Kozikowski - kozikowa@uic.edu; Jia Zhou - Jia.Zhou@psychogenics.com; Joseph H Neale\* - nealej@georgetown.edu

\* Corresponding author

Published: 1 August 2008

Received: 7 May 2008

*Molecular Pain* 2008, **4**:31 doi:10.1186/1744-8069-4-31

Accepted: 1 August 2008

This article is available from: <http://www.molecularpain.com/content/4/1/31>

© 2008 Yamamoto et al; licensee BioMed Central Ltd.

This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/2.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

### Abstract

**Background:** The peptide neurotransmitter *N*-Acetylaspartylglutamate (NAAG) is the third most prevalent transmitter in the mammalian central nervous system. Local, intrathecal and systemic administration of inhibitors of enzymes that inactivate NAAG decrease responses to inflammatory pain in rat models. Consistent with NAAG's activation of group II metabotropic glutamate receptors, this analgesia is blocked by a group II antagonist.

**Results:** This research aimed at determining if analgesia obtained following systemic administration of NAAG peptidase inhibitors is due to NAAG activation of group II mGluRs in brain circuits that mediate perception of inflammatory pain. NAAG and NAAG peptidase inhibitors, ZJ43 and 2-PMPA, were microinjected into a lateral ventricle prior to injection of formalin in the rat footpad. Each treatment reduced the early and late phases of the formalin-induced inflammatory pain response in a dose-dependent manner. The group II mGluR antagonist reversed these analgesic effects consistent with the conclusion that analgesia was mediated by increasing NAAG levels and the peptide's activation of group II receptors.

**Conclusion:** These data contribute to proof of the concept that NAAG peptidase inhibition is a novel therapeutic approach to inflammatory pain and that these inhibitors achieve analgesia by elevating synaptic levels of NAAG within pain processing circuits in brain.

### Introduction

The peptide *N*-acetylaspartylglutamate (NAAG) is by far the most prevalent [1] and widely distributed co-transmitter in the mammalian nervous system [2,3]. It is co-expressed in discrete subsets of neurons with most small amine transmitters, including glutamate and GABA. Consistent with other neuropeptides, NAAG is released under

conditions of high neuronal activity and acts at presynaptic receptors [4-6]. Synaptically released NAAG activates the group II metabotropic glutamate receptors [mGluR3 >> mGluR2; [6-8]]. These receptors are expressed on astrocytes where they stimulate release of trophic factors and on presynaptic axons where they inhibit transmitter release [5,6,9,10]. Two enzymes that inactivate synapti-

cally released NAAG, glutamate carboxypeptidase II and III, have been cloned and characterized [11-15]. Potent inhibitors ( $IC_{50} = 1-5$  nM) of these enzymes are being tested in animal models of neurological conditions that are mediated by high levels of glutamate release [16-18]. While these NAAG peptidase inhibitors do not possess direct agonist activity at ionotropic or metabotropic glutamate receptors, they, like group II mGluR agonists, are effective in reducing perception of inflammatory, neuropathic pain and bone cancer pain in rat models [19-24]. Consistent with the conclusion that inhibitors of NAAG peptidases achieve analgesia by elevating the degree of NAAG activation of a group II mGluR, group II antagonists completely reverse these analgesic actions.

While group II mGluR agonists influence nociceptive responses of primary sensory afferents [19,20,25-28], the widespread distribution of NAAG, NAAG peptidase activity [29] and group II mGluRs within pain pathways (reviewed in [30,31]) suggests that these receptors in the brain also might modulate pain perception following activation by NAAG. Group II mGluRs are upregulated in the central nervous system in response to inflammatory pain states [32-35]. In the periaqueductal grey, a brain region that contributes to descending modulation of nociceptive transmission within the spinal cord [36], group II mGluR agonists act presynaptically to reduce GABAergic transmission [37]. Speculation that this action contributes to analgesia derives from observations that opioid analgesia induced at the level of the periaqueductal grey also is mediated by reduction in GABAergic input to descending projections [38,39]. In this first test of the role of NAAG in regulation of pain perception via brain pain pathways, we administered NAAG and two NAAG peptidase inhibitors into the rat lateral ventricle prior to induction of inflammatory pain.

## Methods

These experiments were executed in adherence with the guidelines of the Committee for Research and Ethical Issues of the International Association for the Study of Pain (1983). They were performed according to a protocol approved by the Institutional Animal Care Committee of Chiba University, Chiba, Japan. Male Sprague-Dawley rats (250 – 300 g, Japan SLC, Shizuoka, Japan) were prepared with ICV catheters and examined for the effect of the agents on the formalin test of inflammatory pain.

### ICV cannulae

Implantation of the intracerebroventricular (ICV) injection cannula into the right lateral ventricle was performed stereotactically under halothane anesthesia. Stainless steel guide cannulae (24 gauge, 0.64 mm outer diameter, 15 mm long) were stereotactically placed through a burr hole (0.5 mm caudal to coronal suture and 1 mm lateral to sag-

ital suture; 3 mm deep to the dura) and affixed to the skull with stainless steel screws and cranioplastic cement. In our experience, drug injection via the cannulae is optimal about 4 days after implantation as the cannulae have not plugged with cells by that time, in contrast to 7 days after implantation. Thus, ICV cannula implantation was performed 4 days before the formalin test. All animals displayed normal feeding and drinking behaviors postoperatively. Rats showing neurological deficits were not studied.

### Formalin test

To carry out the formalin test, 50  $\mu$ l of 5% formalin was injected subcutaneously (SC) into the dorsal surface of the right hind paw with a 25-gauge needle under brief halothane anesthesia. Within 1 min after the formalin injection, spontaneous flinching of the injected paw could be observed. Flinching is readily discriminated and is characterized as a rapid and brief withdrawal or flexion of the injected paw. This pain-related behavior was quantified by counting the number of flinches for 1 min periods at 1 – 2 and at 5 – 6 min, and then for 1 min periods at intervals during the period from 10 to 60 min after the injection. Two phases of spontaneous flinching behavior, an initial acute phase (phase 1: during the first 6 min after the formalin injection) and a prolonged tonic phase (phase 2: beginning about 10 min after the formalin injection), were observed. After the observation period, the animals were immediately killed with an overdose of barbiturate. Four to six rats were used for each treatment group reported here.

### Behavioral analysis

The general behavior of each rat was carefully observed and tested. Motor functions were evaluated by the performance of two specific behavioral tasks, as follows. 1) The placing/stepping reflex: this response was evoked by drawing the dorsum of either hindpaw over the edge of a table top. In normal animals, this stimulus elicits an upward lifting of the paw onto the surface of the table, called stepping. Animals with any degree of hind limb flaccidity will demonstrate an altered or absent reflex. 2) The righting reflex: an animal placed horizontally with its back on the table will normally show an immediate coordinated twisting of the body around its longitudinal axis to regain its normal position on its feet. Animals displaying ataxic behavior will show a decreased ability to right themselves. To quantify the evaluation of motor functions, both tasks were scored on a scale of 0 to 2 in which 0 = absence of function and 2 = normal motor functions. Animals that were able to perform the motor tasks but did so more slowly than normal animals were assigned a score of 1. For example, the reflex withdrawal is typically immediate. Rats demonstrating a non-immediate reflex were scored 1. The normal righting reflex also is prompt and

successful. Rats that either delayed the attempt or who were ultimately but not immediately successful were scored 1.

#### Drugs

(S)-2-[3-[(S)-1-carboxy-3-methylbutyl]ureido]pentanedioic acid (ZJ43) was synthesized following methods previously described [40]. ZJ43 (molecular weight = 304.3) is an unsymmetrical urea, which was prepared by the addition reaction of an isocyanate, generated in situ from the tosylate salt of glutamic acid dibenzyl ester and triphosgene in the presence of  $\text{Et}_3\text{N}$ , with the second amino acid benzyl ester component. The subsequent debenzylation of the key intermediate by the catalytic hydrogenation affords ZJ43 in the final form. 2-(phosphonomethyl) pentanedioic acid (2-PMPA, molecular weight = 314) was purchased from Alexis Biochemicals, (San Diego, CA, USA). ZJ45 and 2-PMPA are NAAG peptidase inhibitors [16-18]. NAAG was purchased from Tocris (Bristol, UK). LY341495, a highly selective group II metabotropic glutamate receptor antagonist [41], was purchased from Tocris. The ICV administered drugs were delivered in a total volume of 3  $\mu\text{l}$ .

#### Experimental protocol

ZJ43, 2-PMPA or NAAG were administered ICV 10 min before the formalin injection. To obtain control data, vehicle (saline) was injected ICV ( $n = 5$ ). To verify that the analgesic effect of ICV administered drugs on the formalin test was mediated by the activation a group II mGluR, 1 mg/kg of LY341495 was administered intraperitoneally (i.p.) 10 min before the ICV injection of drugs. The effect of intraperitoneal (i.p.) administration of 1 mg/kg of LY341495 on the formalin test also was examined.

#### Statistical analyses

For the dose-response analysis, data from phase 1 (0 – 6 min) and phase 2 (10 – 60 min) observations were considered separately. The cumulative instances of formalin-evoked flinches during the phase 1 and phase 2 were calculated for each rat. To evaluate the dose-dependence, one-way analysis of variance (ANOVA) was used. Published data strongly suggest that underlying mechanisms that generate the phase 1 response is different from those that generate the phase 2 response. Thus, for the dose-response analysis, we considered phase 1 and phase 2 responses as independent samples. For multiple comparisons, Tukey's test was used. For comparison of the pain response in the peptidase inhibitor group versus the group that was treated with inhibitor and the group II antagonist, the unpaired t-test (two tailed) was used.

Whenever appropriate, results are expressed as mean  $\pm$  SEM. Critical values that reached a  $p < 0.05$  level of significance were considered significant.

## Results

### Behavioral analysis

ICV injection of 100  $\mu\text{g}$  of ZJ43, caused restlessness in all animals and 20% of the rats receiving 100  $\mu\text{g}$  of ZJ43 scored 1 (impairment of motor function) in the placing/stepping reflex and righting reflex. After ICV injection of 10  $\mu\text{g}$  or less of NAAG, ZJ43 or 2-PMPA, all animals scored 2 (normal motor function) in the placing/stepping reflex and righting reflex tests. As a result, 10  $\mu\text{g}$  was the highest dose of ZJ43 and 2-PMPA injected into the lateral ventricle in this study. After the i.p. administration of 1 mg/kg of LY341495, all animals scored 2 (normal motor function) in the placing/stepping reflex and righting reflex tests.

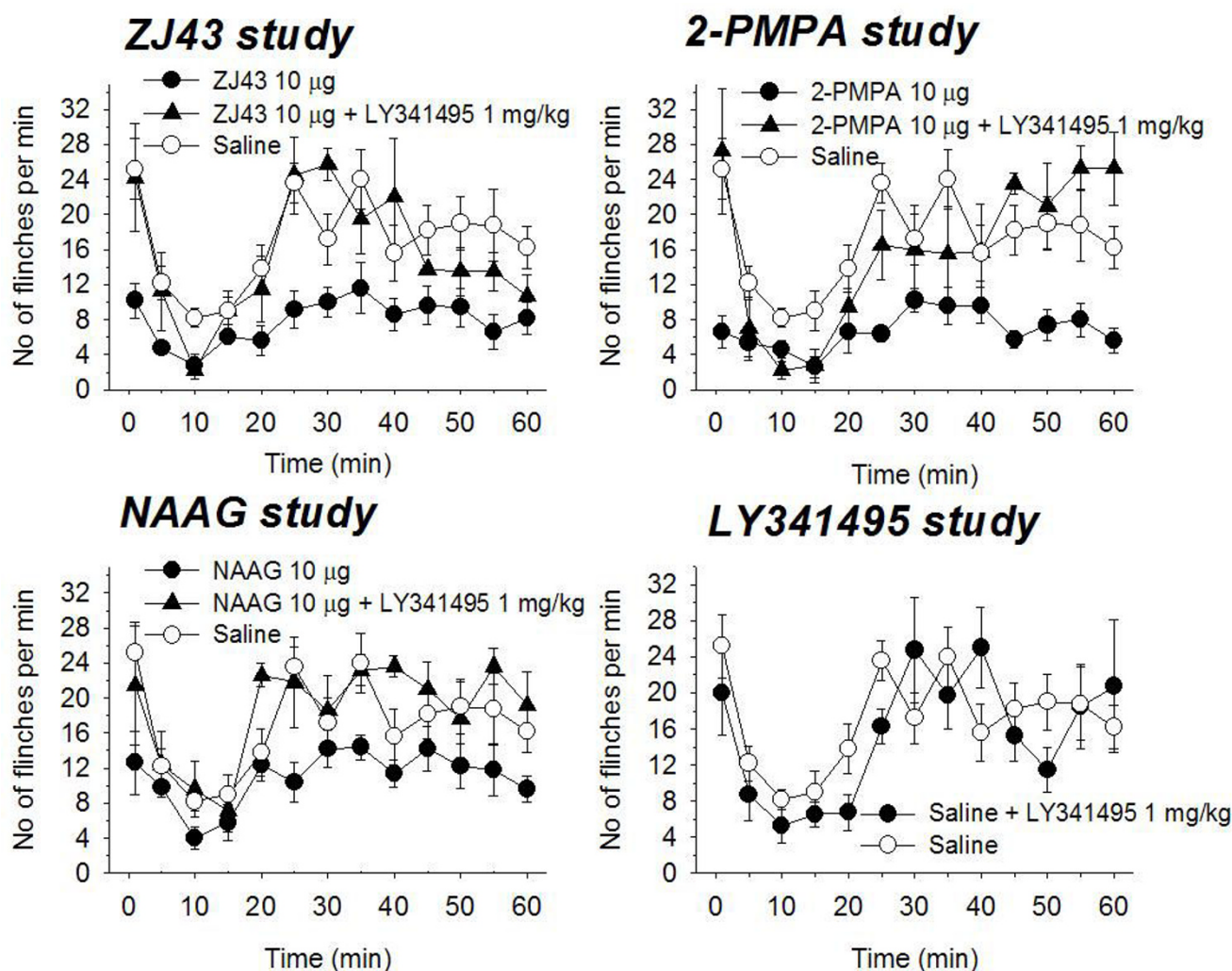
### Responses in formalin model of inflammatory pain

ICV injection of ZJ43 (Figure 1a), 2-PMPA (Figure 1b) or NAAG (Figure 1c) decreased the sum of flinches induced by formalin injection into the footpad while the group II mGluR agonist LY341495 alone (Figure 1c) had no detectable effect. Pretreatment with the group II mGluR antagonist LY341495 (1 mg/kg, i.p.) blocked the analgesic effect of 10  $\mu\text{g}$  of ZJ43 or 2-PMPA on both phases of the flinching behavior (Figures 1a and 1b; phase 1:  $p < 0.01$ ; phase 2:  $p < 0.01$ , by t-test). LY341495 similarly antagonized the analgesic effect of 10  $\mu\text{g}$  of NAAG on the phase 2, but not phase 1, flinching behavior (Figure 1c; phase 1:  $p > 0.2$ ; phase 2:  $p < 0.005$ , by t-test).

The NAAG peptidase inhibitors reduced phase 1 and phase 2 flinching behaviors in a dose-dependent manner relative to saline treated rats (Figures 2a and 2b; phase 1:  $p < 0.01$ ; phase 2:  $p < 0.01$  by ANOVA). Similarly, ICV injection of NAAG itself decreased the sum of flinches, in both phases of the flinching behavior in a dose-dependent manner between 1 and 10  $\mu\text{g}$  (Figures 2a and 2b; phase 1:  $p < 0.05$ ; phase 2:  $p < 0.01$  by ANOVA). The maximal effect of the peptide appeared to be less than that obtained by the peptidase inhibitors. Ten  $\mu\text{g}$  of NAAG provided a maximal effect inasmuch as 100  $\mu\text{g}$  of NAAG gave no greater reduction (Figure 2a and 2b).

## Discussion

We previously reported that systemic, intrathecal and local application of NAAG peptidase inhibitors reduced pain perception in rat models of inflammatory, neuropathic and bone cancer induced pain and that these effects are blocked by co-application of a group II mGluR [20-24]. These and other data support the conclusion that the analgesic effects of NAAG peptidase inhibition are mediated by increased activation of presynaptic group II mGluRs and a subsequent reduction in transmitter release in those neuronal circuits in which NAAG and the peptidase activity are expressed [reviewed in [16]]. However, it was not clear from these data if systemic application of

**Figure 1**

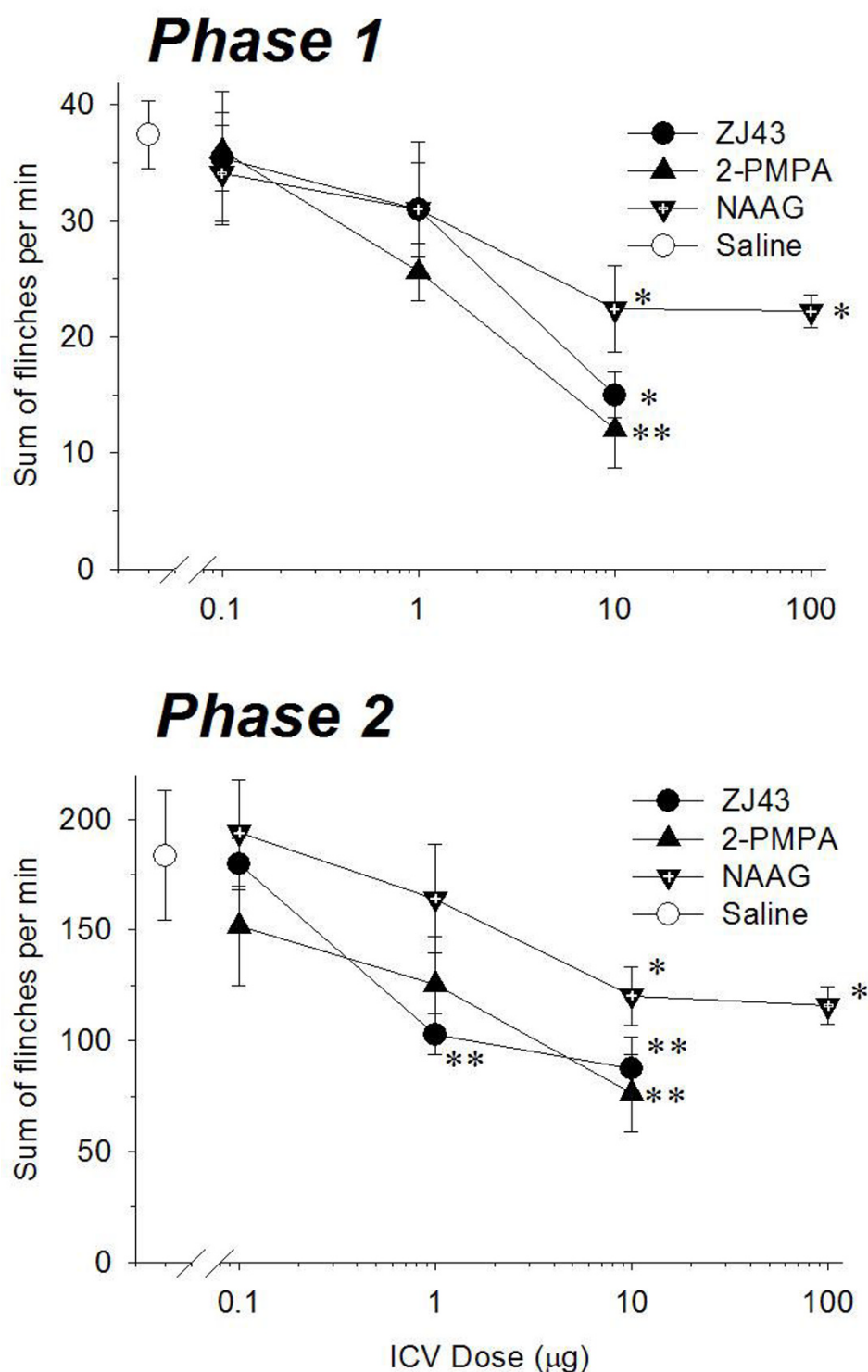
**Effects of intracerebroventricular (ICV) injection of 10 µg of ZJ43 (a), 2-PMPA (b) or NAAG (c) on the time course of the flinches observed after the formalin injection into the dorsal surface of the right rat hind-paw.**

Drugs were administered 10 min before the formalin injection. The group II mGluR antagonist LY341495 (1 mg/kg, i.p.) was injected 10 min before ZJ43, 2-PMPA, NAAG or saline (d). The number of flinches/min is plotted vs time after the formalin injection. NAAG and the NAAG peptidase inhibitors ZJ43 and 2-PMPA significantly reduced both phases of the response to formalin injection (see text for statistical assessment). The effects of the peptidase inhibitors were blocked by the LY341495 as was the effect of NAAG on phase 2 of the response to formalin. Each line represents the group mean and S.E.M. of four to six rats.

NAAG peptidase inhibitors achieved analgesia by activation of group II mGluRs in brain pain pathways. This research ultimately is aimed at testing the concept that NAAG peptidase inhibition represents a clinically significant and completely new strategy for the treatment of inflammatory and neuropathic pain. As such, defining the locus of action of these peptidase inhibitors relative to pain perception is an issue of central importance. The data presented here support the conclusion that while the spinal cord and sensory neurons represent loci at which NAAG peptidase inhibition may contribute to analgesia,

there also is at least one locus in the brain where NAAG directly mediates analgesia and that this brain region is accessible via the lateral ventricles. Given the quantities of the ZJ43 and 2-PMPA that are required locally [20] and intrathecally [21] to obtain analgesia, it is possible that the primary target for systemically applied NAAG peptidase inhibitor-mediated analgesia is in the brain and/or brain stem.

Since the inhibitors were infused into the right ventricle and the inflammation was induced in the ipsilateral foot-

**Figure 2**

**Dose-response curves for ICV injection of ZJ43, 2-PMPA and NAAG representing the cumulative instances of formalin evoked flinches during the phase I (a) and the phase 2 (b).** ZJ43, 2-PMPA and NAAG reduced the number of phase I and the phase 2 flinching behaviors in a dose dependent manner. Each point represents the group mean and S.E.M. of responses by groups of 5–6 rats. \* <0.05 and \*\* <0.005 versus rats given saline ICV prior to formalin-induced inflammation.

pad, it seems likely that the inhibitors were not acting proximal to this ventricle but rather that the compounds were affecting the broader periventricular tissue of the brain including the contralateral descending pain modulating pathway in the periaqueductal grey. The periaqueductal grey is an important locus in the pathway for opioid mediated analgesia. Relevant to these NAAG peptidase inhibition data, the periaqueductal grey contains a relatively high concentration of both NAAG and NAAG peptidase activity [29]. Further, group II mGluR agonist activation of presynaptic receptors on GABAergic neurons in the periaqueductal grey reduces GABA release [37]. We previously reported that NAAG inhibits GABA release from cortical neurons via a presynaptic mechanism [5]. A similar action of presynaptic opiate receptors on GABAergic neurons in the periaqueductal grey disinhibits a descending pathway that suppresses nociceptive transmission in the dorsal horn of the spinal cord [36,39]. While these data are consistent with speculation that intracerebroventricular administration of NAAG peptidase inhibitors might achieve analgesia via a mechanism that parallels that of the opiates in the PAG, a dose response study directly in the PAG is required to demonstrate this site of action. More importantly, these data together with our previous studies suggest that this peptide transmitter functions both spinally and centrally along pathways that mediate and modulate perception of inflammatory pain in a manner that is analogous to opiate peptides.

Given the wide distribution of group II mGluR receptors in the brain, it is surprising that direct infusion of NAAG did not result in a broad range of behavioral effects outside of the pain modulatory pathway and that its effects on the pain responses were not at least as profound as those of the peptidase inhibitors. This result is most likely due to the equally widespread distribution of extracellular NAAG peptidase activity in the brain. In the absence of peptidase inhibition, NAAG delivered by ICV is likely to be hydrolyzed rapidly as it diffuses through the extracellular space. Equally relevant to the development of new analgesic drugs, the systemic administration of the peptide would have the same drawbacks as synthetic group II agonists. That is, agonists activate receptors without a relationship to the normal level of information movement within circuits. In contrast, strategies that enhance the actions of endogenously released transmitters, such as NAAG peptidase inhibition, enhance the effects of the normal action of the circuit. This is, for example, the basis of the therapeutic efficacy of benzodiazepines, barbiturates and serotonin selective reuptake inhibitors.

ZJ43 is tricarboxylic acid with a urea core. Its hydrophilic structure contributes to its relatively low penetration across the CACO-2 cell culture model of tight junctions as found in the blood-brain barrier (Neale, et al., unpub-

lished data). As a result, it is not surprising that despite its low nanomolar  $IC_{50}$  for NAAG peptidases, relatively high (50–100 mg/kg) systemic doses of ZJ43 are required to obtain significant analgesia in inflammatory and neuropathic pain models [21]. This finding that some of the analgesic effects of ZJ43 are obtained by acting at pain circuits that are accessible via the lateral ventricle militates in favor of developing similarly potent NAAG peptidase inhibitors that cross the blood brain barrier more effectively. Toward this objective, we have synthesized a series of prodrug esters of ZJ43 that are themselves not NAAG peptidase inhibitors but that may enter the central nervous system more effectively due to their improved ClogP values. In preliminary studies, some of these prodrugs are more active in an inflammatory pain model than is ZJ43 (Yamamoto, unpublished). These results suggest that some esters of ZJ43 penetrate the blood-brain barrier effectively and are hydrolyzed efficiently by central nervous system esterases rather than by serum esterases. If NAAG peptidase inhibition continues to show promise as an analgesic pharmacotherapy, this prodrug development strategy is likely to be important in reducing the amount of drug required to achieve analgesia, thus reducing the probability of secondary effects and toxicity in other tissues.

## Conclusion

NAAG and its receptors are present in circuits within the brain and brainstem that process pain perception. This study provides the first direct demonstration that NAAG peptidase inhibitors reduce inflammatory pain perception by acting on pain communication pathways within the brain. Currently, clinical therapy for pain is limited to opioids and non-steroidal analgesics. This discovery of the analgesic effects of NAAG and NAAG peptidase inhibitors in the brain represents an important step in understanding the cellular mechanism underlying this promising new approach to analgesia.

## Abbreviations

N-acetylaspartylglutamate: NAAG; metabotropic glutamate receptor: mGluR.

## Competing interests

Tatsuo Yamamoto and Joseph Neale have no competing interests. This work was completed during an interval when Acenta Discovery held the license from Georgetown University to the intellectual property related to ZJ43. Dr. Kozikowski was the major owner of Acenta Discovery, Inc. Dr. Zhou was an employee of Acenta Discovery. Acenta Discovery is now owned by PsychoGenics, Inc., and Dr. Zhou is now an employee of PsychoGenics Inc. As such, Dr. Kozikowski and Dr. Zhou could profit from this class of compounds should they ever become drugs. Neither



had any direct involvement in collection or analysis of data presented herein.

### Authors' contributions

TY executed the behavioral studies. JHN worked with AK and JZ on the characterization of ZJ43, generated the hypothesis, created the experimental design and wrote the manuscript. AK and JZ intellectual contribution was the designed ZJ43 within a series of related NAAG peptidase inhibitors. Additionally, JZ synthesized and chemically characterized ZJ43. All authors have reviewed and approved this final manuscript.

### Acknowledgements

This research supported by a grant from the NIH (NS38080) to JHN, Grant-in-Aid for Scientific Research (B) of Japan (12470315) to TY and gifts to Georgetown University by Nancy and Daniel Paduano (JHN). Acenta Discovery, Inc. supported this research by providing ZJ43.

### References

- Curatolo A, D'Archangelo P, Lino A, Brancati A: **Distribution of N-acetyl-aspartic and N-acetyl-aspartyl-alutamic acids in nervous tissue.** *J Neurochem* 1965, **12**:339-342.
- Coyle JT: **The nagging question of the function of N-acetylaspartylglutamate.** *Neurobiol Dis* 1997, **4**:231-238.
- Neale JH, Bzduga T, Wroblewska B: **N-Acetylaspartylglutamate: the most abundant peptide neurotransmitter in the mammalian central nervous system.** *J Neurochem* 2000, **75**:443-452.
- Zhong C, Zhao X, Sarva J, Kozikowski A, Neale JH, Lyeth BG: **NAAG peptidase inhibitor reduces acute neuronal degeneration and astrocyte damage following lateral fluid percussion TBI in rats.** *J Neurotrauma* 2005, **22**:266-276.
- Zhao J, Ramadan E, Capiello M, Wroblewska B, Bzduga T, Neale JH: **NAAG inhibits KCl-induced [(3)H]-GABA release via mGluR3, cAMP, PKA and L-type calcium conductance.** *Eur J Neurosci* 2001, **13**:340-346.
- Sanabria ER, Wozniak KM, Slusher BS, Keller A: **GCP II (NAALADase) inhibition suppresses mossy fiber-CA3 synaptic neurotransmission by a presynaptic mechanism.** *J Neurophysiol* 2004, **91**:182-193.
- Wroblewska B, Wroblewski JT, Pshenichkin S, Surin A, Sullivan SE, Neale JH: **N-acetylaspartylglutamate selectively activates mGluR3 receptors in transfected cells.** *J Neurochem* 1997, **69**:174-181.
- Wroblewska B, Santi MR, Neale JH: **N-acetylaspartylglutamate activates cyclic AMP-coupled metabotropic glutamate receptors in cerebellar astrocytes.** *Glia* 1998, **24**:172-179.
- Bruno V, Battaglia G, Casabona G, Copani A, Caciagli F, Nicoletti F: **Neuroprotection by glial metabotropic glutamate receptors is mediated by transforming growth factor-beta.** *J Neurosci* 1998, **18**:9594-9600.
- Vignes M, Clarke VR, Davies CH, Chambers A, Jane DE, Watkins JC, Collingridge GL: **Pharmacological evidence for an involvement of group II and group III mGluRs in the presynaptic regulation of excitatory synaptic responses in the CA1 region of rat hippocampal slices.** *Neuropharmacology* 1995, **34**:973-982.
- Israeli RS, Powell CT, Fair VWR, Heston VWD: **Molecular cloning of a complementary DNA encoding a prostate-specific membrane antigen.** *Cancer Res* 1993, **53**:227-230.
- Carter RE, Feldman AR, Coyle JT: **Prostate-specific membrane antigen is a hydrolase with substrate and pharmacologic characteristics of a neuropeptidase.** *Proc Natl Acad Sci USA* 1996, **93**:749-753.
- Bzduga T, Turi T, Wroblewska B, She D, Chung HS, Kim H, Neale JH: **Molecular cloning of a peptidase against N-acetylaspartylglutamate from a rat hippocampal cDNA library.** *J Neurochem* 1997, **69**:2270-2277.
- Luthi-Carter R, Berger UV, Barczak AK, Enna M, Coyle JT: **Isolation and expression of a rat brain cDNA encoding glutamate carboxypeptidase II.** *Proc Natl Acad Sci USA* 1998, **95**:3215-3220.
- Bzduga T, Crowe SL, Ramadan ER, Sciarretta KH, Olszewski RT, Ojeifo OA, Rafalski VA, Wroblewska B, Neale JH: **The cloning and characterization of a second brain enzyme with NAAG peptidase activity.** *J Neurochem* 2004, **89**:627-635.
- Neale JH, Olszewski RT, Gehl LM, Wroblewska B, Bzduga T: **The neurotransmitter N-acetylaspartylglutamate in models of pain, ALS, diabetic neuropathy, CNS injury and schizophrenia.** *Trends Pharmacol Sci* 2005, **26**:477-484.
- Zhou J, Neale JH, Pomper MG, Kozikowski AP: **NAAG peptidase inhibitors and their potential for diagnosis and therapy.** *Nat Rev Drug Discov* 2005, **4**:1015-1026.
- Tsukamoto T, Wozniak KM, Slusher BS: **Progress in the discovery and development of glutamate carboxypeptidase II inhibitors.** *Drug Discov Today* 2007, **12**:767-776.
- Sharpe EF, Kingston AE, Lodge D, Monn JA, Headley PM: **Systemic pre-treatment with a group II mGlu agonist, LY37 reduces hyperalgesia in vivo.** *Br J Pharmacol* 1996, **117**:1255-1262.
- Yamamoto T, Saito O, Aoe T, Bartolozzi A, Sarva J, Zhou J, Kozikowski A, Wroblewska B, Bzduga T, Neale JH: **Local administration of N-acetylaspartylglutamate (NAAG) peptidase inhibitors is analgesic in peripheral pain in rats.** *Eur J Neurosci* 2007, **25**:147-158.
- Yamamoto T, Hirasawa S, Wroblewska B, Grajkowska E, Zhou J, Kozikowski A, Wroblewski J, Neale JH: **Antinociceptive effects of N-acetylaspartylglutamate (NAAG) peptidase inhibitors ZJ-11, ZJ-17 and ZJ-43 in the rat formalin test and in the rat neuropathic pain model.** *Eur J Neurosci* 2004, **20**:483-494.
- Yamamoto T, Nozaki-Taguchi N, Sakashita Y, Inagaki T: **Inhibition of spinal N-acetylated-alpha-linked acidic dipeptidase produces an antinociceptive effect in the rat formalin test.** *Neuroscience* 2001, **102**:473-479.
- Yamamoto T, Nozaki-Taguchi N, Sakashita Y: **Spinal N-acetyl-alpha-linked acidic dipeptidase (NAALADase) inhibition attenuates mechanical allodynia induced by paw carrageenan injection in the rat.** *Brain Res* 2001, **909**:138-144.
- Saito O, Aoe T, Kozikowski A, Sarva J, Neale JH, Yamamoto T: **Ketamine and N-acetylaspartylglutamate peptidase inhibitor exert analgesia in bone cancer pain.** *Can J Anaesth* 2006, **53**:891-898.
- Yang D, Gereau RWt: **Peripheral group II metabotropic glutamate receptors (mGluR2/3) regulate prostaglandin E2-mediated sensitization of capsaicin responses and thermal nociception.** *J Neurosci* 2002, **22**:6388-6393.
- Yang D, Gereau RWt: **Peripheral group II metabotropic glutamate receptors mediate endogenous anti-allodynia in inflammation.** *Pain* 2003, **106**:411-417.
- Yang D, Gereau RWt: **Group II metabotropic glutamate receptors inhibit cAMP-dependent protein kinase-mediated enhancement of tetrodotoxin-resistant sodium currents in mouse dorsal root ganglion neurons.** *Neurosci Lett* 2004, **357**:159-162.
- Ahn DK, Kim KH, Jung CY, Choi HS, Lim EJ, Youn DH, Bae YC: **Role of peripheral group I and II metabotropic glutamate receptors in IL-1beta-induced mechanical allodynia in the orofacial area of conscious rats.** *Pain* 2005, **118**:53-60.
- Fuhrman S, Palkovits M, Cassidy M, Neale JH: **The regional distribution of N-acetylaspartylglutamate (NAAG) and peptidase activity against NAAG in the rat nervous system.** *J Neurochem* 1994, **62**:275-281.
- Neugebauer V: **Metabotropic glutamate receptors - important modulators of nociception and pain behavior.** *Pain* 2002, **98**:1-8.
- Varney MA, Gereau RWt: **Metabotropic glutamate receptor involvement in models of acute and persistent pain: prospects for the development of novel analgesics.** *Curr Drug Targets CNS Neurol Disord* 2002, **1**:283-296.
- Neto FL, Schadrack J, Platzer S, Ziegler-Schaber W, Tolle TR, Castro-Lopes JM: **Up-regulation of metabotropic glutamate receptor 3 mRNA expression in the cerebral cortex of monoarthritic rats.** *J Neurosci Res* 2001, **63**:356-367.
- Dolan S, Kelly JG, Monteiro AM, Nolan AM: **Up-regulation of metabotropic glutamate receptor subtypes 3 and 5 in spinal cord in a clinical model of persistent inflammation and hyperalgesia.** *Pain* 2003, **106**:501-512.
- Dolan S, Kelly JG, Monteiro AM, Nolan AM: **Differential expression of central metabotropic glutamate receptor (mGluR)**

- subtypes in a clinical model of post-surgical pain. *Pain* 2004, **110**:369-377.
35. Han JS, Fu Y, Bird GC, Neugebauer V: **Enhanced group II mGluR-mediated inhibition of pain-related synaptic plasticity in the amygdala.** *Mol Pain* 2006, **2**:18.
  36. Fields HL, Basbaum AI: **Central nervous system mechanisms of pain modulation.** In *Textbook of Pain* Edited by: Wall PD, Melzack R. Textbook of Pain, Edinburgh: Churchill Livingstone; 1999:309-322.
  37. Drew GM, Vaughan CW: **Multiple metabotropic glutamate receptor subtypes modulate GABAergic neurotransmission in rat periaqueductal grey neurons in vitro.** *Neuropharmacology* 2004, **46**:927-934.
  38. Osborne PB, Vaughan CW, Wilson HI, Christie MJ: **Opioid inhibition of rat periaqueductal grey neurones with identified projections to rostral ventromedial medulla in vitro.** *J Physiol* 1996, **490(Pt 2)**:383-389.
  39. Vaughan CW, Christie MJ: **Presynaptic inhibitory action of opioids on synaptic transmission in the rat periaqueductal grey in vitro.** *J Physiol* 1997, **498(Pt 2)**:463-472.
  40. Kozikowski AP, Zhang J, Nan F, Petukhov PA, Grajkowska E, Wroblewski JT, Yamamoto T, Bzdega T, Wroblewska B, Neale JH: **Synthesis of urea-based inhibitors as active site probes of glutamate carboxypeptidase II: efficacy as analgesic agents.** *J Med Chem* 2004, **47**:1729-1738.
  41. Kingston AE, Ornstein PL, Wright RA, Johnson BG, Mayne NG, Burnett JP, Belagaje R, Wu S, Schoepp DD: **LY341495 is a nanomolar potent and selective antagonist of group II metabotropic glutamate receptors.** *Neuropharmacology* 1998, **37**:1-12.

Publish with **BioMed Central** and every scientist can read your work free of charge

"BioMed Central will be the most significant development for disseminating the results of biomedical research in our lifetime."

Sir Paul Nurse, Cancer Research UK

Your research papers will be:

- available free of charge to the entire biomedical community
- peer reviewed and published immediately upon acceptance
- cited in PubMed and archived on PubMed Central
- yours — you keep the copyright

Submit your manuscript here:  
[http://www.biomedcentral.com/info/publishing\\_adv.asp](http://www.biomedcentral.com/info/publishing_adv.asp)

